



Review Article

Glycation End-Products and their Receptors: Pathophysiology and Therapeutic Targeting in Diabetes Mellitus

Hussain Saad Abdulrahman^{1*}, Naza Mohammed Ali Mahmood²

¹ Department of Pharmacy, Shaheed Salah Teaching Hospital, Sulaimani, Kurdistan Region, Iraq

² Department of Pharmacology and Toxicology, College of Pharmacy, University of Sulaimani, Kurdistan Region, Iraq

Received: June 2021; Revised: July 2021; Accepted: August 2021

Abstract

Diabetes mellitus (DM) impairs cell metabolism and function in a variety of organs, increasing the risk of pathologies in organs such the kidney, neurological system, and eye, as well as fragility fractures. Advanced glycation end products (AGEs) are chemical moieties created by long-term hyperglycemia that interact with specific AGE receptors (RAGEs) to affect cellular metabolism and/or function. Some of the clinical effects of DM on cellular metabolism and organ function through the AGE-RAGE signaling pathway were detected through PubMed searches using the keywords "advanced glycation end product "RAGE", "sRAGE", "DM", and "complications." Tables were created for all published experimental and clinical research. Diabetic sequelae such as nephropathy, neuropathy, retinopathy, and osteopathy are all linked to AGE-RAGE signaling. Some clinical outcomes in diabetic individuals could be attributable to the effects of AGE-RAGE signaling. The AGE-RAGE signaling system, on the other hand, has some beneficial effects in a variety of tissues, including an increase in osteogenic function. As a ligand decoy, soluble RAGE (sRAGE) may increase in either RAGE production or destruction, and it cannot always reflect AGE-RAGE signaling. The AGE-RAGE axis can be targeted using a variety of drugs. They can also mitigate the negative outcomes. Although recombinant sRAGE can block the AGE-RAGE signaling pathway, it has numerous drawbacks, including AGE accessibility, an increase in other RAGE ligands, and a lengthy half-life (24 hours). It's linked to the loss of AGE/positive RAGE's effects. As a result, sRAGE is not a useful marker for assessing the RAGE signaling pathway's activation. Due to its limitations, recombinant sRAGE cannot be used in clinical practice.

Keywords: AGEs, Diabetes Mellitus, DM complications, RAGEs

النواتج النهائية لتفاعل الجلوزة ومستقبلاتها: الفيزيولوجيا المرضية والاستهداف العلاجي في علاج مرض السكري

الخلاصة

يضعف داء السكري استقلاب الخلايا والوظائف في مختلف أعضاء الجسم، مما يزيد من خطر اعتلال أعضاء مثل الكلى والجهاز العصبي والعين، فضلاً عن هشاشة العظام. المنتجات النهائية المتقدمة للجليكة هي مركبات كيميائية تنتج عن فرط السكر في الدم على المدى الطويل والتي تتفاعل مع مستقبلات محددة للتأثير على وظائف الخلايا. تم الكشف عن بعض الآثار السريرية لداء السكري باستخدام الإشارات AGE-RAGE خلال عمليات البحث في PubMed وباستخدام الكلمات الرئيسية "المنتج النهائي لعملية التصلب الجانبي المتقدمة و"المضاعفات". وتم إنشاء جداول لجميع البحوث التجريبية والسريرية المنشورة حول الموضوع. من تبعات السكري اعتلال الكلى، والاعتلال العصبي، اعتلال الشبكية، واعتلال العظام، وكلها مرتبطة بإشارات AGE-RAGE. يمكن أن تعزى بعض النتائج السريرية في الأفراد المصابين بالسكري إلى تأثير ارتباط نواتج الجلوزة مع مستقبلاتها، من ناحية أخرى، لديه بعض الآثار المفيدة في مجموعة متنوعة من الأنسجة، بما في ذلك زيادة في وظيفة العظام. يمكن استهداف محور AGE-RAGE باستخدام مجموعة متنوعة من الأدوية. على الرغم من أن المستقبل الذائب يمكن أن يمنع مسار الإشارة AGE-RAGE، إلا أنه يحتوي على العديد من العيوب. ونتيجة لذلك، فإنها ليست مفيدة لتقييم نشاط مسار هذا الارتباط، وعليه لا يمكن استخدام المستقبل المؤتلف في التجارب السريرية.

* **Corresponding author:** Hussain S. Abdulrahman; Department of Pharmacy, Shaheed Salah Teaching Hospital, Sulaimani, Kurdistan Region, Iraq; Email: hussein.alzadi90@gmail.com

Article citation: Abdulrahman HS, Mahmood NMA. Glycation end-products and their receptors: Pathophysiology and therapeutic targeting in diabetes mellitus. *Al-Rafidain J Med Sci.* 2021;1:19-35.

INTRODUCTION

Fast food, which is high in carbohydrates and calories, and canned food, which has many additives such as colors, flavors, and taste, are both influenced by modern lifestyles. Elevated plasma glucose levels are caused by consuming such kind of diets [1]. As a result, metabolic illnesses including diabetes and obesity are more common in developed and industrialized countries. Diabetes Mellitus (DM) is a category of metabolic illnesses caused by insulin insufficiency (T1DM), insulin resistance (T2DM), or a combination of both. Its complications represent a global burden in terms of both health and economics [2]. Over the previous few decades, the rate of global propagation has increased rapidly. According to the International Diabetes Federation (IDF), there are approximately 400 million people worldwide aged 18 to 98, and the number of adult diabetic patients is expected to rise in the next decades. Adults diagnosed with diabetes are anticipated to number more than 650 million by 2045, due to a variety of factors including poor fast food consumption, lifestyle changes, lack of physical activity, and urbanization [3,4]. Diabetes Mellitus claimed the lives of around 1.6 million individuals worldwide in 2015 [5]. DM has a significant morbidity rate due to the vast spectrum of catastrophic consequences. Many organs and tissues are affected by chronic hyperglycemia, which is caused by a lack of insulin or insulin resistance. It causes major side effects such as nephropathy, neuropathy, retinopathy, and, most notably, cardiovascular events [6,7]. Increased protein glycation and gradual elevation of advanced glycated end products (AGES) in the tissues are caused by hyperglycemia. They were important in the development of DM Complications [8]. Diabetic people are four times more likely than non-diabetic persons to develop peripheral vascular diseases [9].

GLYCATION REACTION

The reaction between the carbonyl group of carbohydrates and an amino acid group of various substances such as protein, DNA, and lipids is known as glycation. The glycation process can take place in one of two ways: The first is enzymatic-dependent glycation, such as glycoprotein formation, while the second is non-enzymatic-dependent glycation (glycosylation), such as the chemical interaction between reducing sugar and proteins, as shown by the reaction of glucose with protein lysine residues to create ketoamine (Amadori adduct). The Maillard reaction is a non-enzyme-dependent reaction between the reducing sugar and amino acid terminus of proteins that occurs frequently during food processing. In the food, these complicated processes result in the

creation of a brown products [10]. There are three stages of the non-enzymatic glycation process: initial, middle, and final [11]. The glycation process produces reversible unstable Schiff base through a covalent link between carbonyl and amino residue in the first stage, at a high temperature. The latter undergoes an Amadori reaction, which results in the formation of Amadori rearrangement products (ARPs), which are colorless and UV-insensitive [12]. ARPs reactions are pH-dependent reactions in the intermediate stage [11]. When ARPs are converted to furfural and hydroxymethyl furfural at pH 7, they mostly progress (HMF). Meanwhile, ARPs are converted to dehydroreductones and reductones at pH > 7 and low temperatures, resulting in the synthesis of aldehydes and aminoketones. After UV absorption, the color of the products changed to yellow at this stage. When the pH is greater than 7 and the temperature is high, additional compounds such as glyoxal (GO), methylglyoxal (MG), and diacetyl derivatives are generated. After UV absorption, the color of the products may turn yellow [13]. Many chemical reactions, such as condensation, dehydrogenation, isomerization, and rearrangement, create brownish low molecular weight (LMW) and high molecular weight (HMW) nitrogen polymers in the end stage. Melanoidins are the end products of browning without the need of enzymes. They are not to be confused with melanins, which are the end products of enzymatic activities [11]. The glycation process and the generation of glycation end-products happened at the same time [12]. Fournet *et al.* showed that the glycation process played a role in the pathophysiology of age-related non-communicable chronic illnesses affecting numerous organs [14].

ADVANCED GLYCATION END PRODUCTS (AGES)

Advanced glycation end products (AGEs), also known as glycotoxins, is a broad word that refers to the molecules formed when a reducing sugar reacts with amino acid residues in proteins, lipids, and nucleic acids without the assistance of enzymes (Maillard reaction). Furthermore, excessive AGEs production might trigger and increase the process of oxidative stress within cells [15]. Furthermore, various processes, including the oxidation of carbohydrates, lipids, and amino acids, can produce reactive aldehydes that covalently bond to proteins, resulting in AGEs formation. On the other hand, a high amount of reactive oxygen species (ROS) encourages the synthesis and accumulation of AGEs. Many *in vivo* investigations have shown that oxidative stress caused by AGEs overproduction causes a variety of brain damages that can lead to illnesses [16]. AGEs can be

produced exogenously or endogenously. Exogenous AGEs can be produced by consuming roasted or broiled foods at high temperatures, as well as processed foods. The non-enzymatic glycation reaction is aided by the higher temperature (Maillard reaction). This reaction occurs when the carbonyl group of a reducing sugar and the amino group of a protein combine to generate an unstable reversible compound. The resulted Schiff's base is subsequently subjected to the Amadori rearrangement procedure, which produces Amadori rearrangement products (ARPs). The pH and temperature of the rearrangement stage are also important. Finally, ARPs undergo a series of complicated processes such as condensation and isomerization, which result in the formation of AGEs [17]. Many different types of AGEs have been identified, including 3DG-H1, 3-deoxyglucosone-derived hydroimidazolone 1, N-carboxyethyl-arginine (CEA), N-carboxyethyl-lysine (CEL), N-carboxymethyl-arginine (CMA), carboxymethyl cysteine (CMC), N-carboxymethyl-lysine (CML), 3-deoxyglucosone-2-oxoethyl-lysine, glyoxal-derived lysine dimer (GOLD), HbA1c, methylglyoxal-derived hydroimidazolone 1 (MG-H1), methylglyoxal-derived hydroimidazolone 3 (MG-H3), methylglyoxal-derived imidazolium crosslink (MODIC), and methylglyoxal-derived lysine dimer (MOLD). These chemicals are utilized in food processing because they have unique qualities (aroma, taste, and flavor). They will be absorbed from the intestine and reach circulation when high AGEs containing foods are consumed. They have a negative impact on a variety of organs [20]. Furthermore, AGEs are eliminated in the urine within 48 hours, and high serum levels of AGEs have been documented in renal failure patients [21]. Endogenous AGEs can also be formed by endogenous non-enzyme-dependent glycation of lipids and proteins [22], which is aided by chronic illnesses including hypertension and diabetes [23]. The accumulation of AGEs has been shown to be a biomarker of aging and has been linked to poor outcomes in both DM therapy and surgical interventions [24]. The production and actions of AGEs are governed by two processes. The first functions by using an internal glycosylase system (glyoxalase I and II) to prevent the synthesis of dicarbonyl molecules [25]. The interaction of AGEs with particular receptors is the second process (sRAGE, esRAGE). As a result of the lack of an intracellular signal, such interactions inhibit the effects of AGEs [26]. The action of cellular enzymes and the antiglycation mechanism are involved in the repair of damaged proteins protect cells and tissues in healthy persons against the formation of AGEs and other hazardous substances [27]. Our bodies, on the other

hand, have no control over the formation and accumulation of AGEs during illnesses [28]. Furthermore, Eisermann *et al.* found that the ubiquitin-proteasome system (UPS) and autophagy are two other mechanisms that contribute to the detection and elimination of AGEs in human bodies [29].

AGEs RECEPTORS (RAGEs)

AGEs aren't just thought of as indicators for aging, hyperglycemia, inflammation, and oxidative stress. Many pathophysiological disorders are caused by the interaction of AGEs with their receptors (RAGEs). Not all AGEs have the same affinity for RAGEs; nonetheless, methylglyoxal has a significant (high affinity) interaction with RAGEs [30]. RAGE is a transmembrane receptor for advanced glycation end products. It has an intracellular tail and a highly hydrophobic transmembrane domain, and an extracellular region with three immunoglobulin-like domains, one V-type and two C-type (C1 and C2) domains [31]. The V-type domain is required for ligand binding. RAGEs are now identified as soluble forms (sRAGE) and can be found in a variety of biological fluids, including plasma, synovial fluid, CSF, and bronchoalveolar fluid [32,33]. sRAGE isoforms include sRAGE1/2/3, esRAGE (endogenous soluble RAGE), and hRAGEsec, among others (human RAGE secreted). The synthesis of distinct sRAGE isoforms is attributed to alternative splicing and proteolytic cleavage processes, according to numerous studies [34]. The majority of circulating sRAGEs were produced by splitting off the full-length receptor's cell surface. Matrix metalloproteinases (MMPs) and disintegrin are primarily responsible for this expression [35]. The synthesis of sRAGE is also linked to the G-protein coupled receptor [36]. Meanwhile, esRAGE is a less common variant of RAGEs [37] that results from alternative splicing of a variant RAGE form. Other than AGEs, RAGE binds to a variety of ligands, including high mobility group protein B1, S100 calcium-binding proteins (e.g., calgranulin), amyloid protein, and amphotericin [38]. Furthermore, AGEs can bind to a variety of receptors without triggering intracellular signaling, as evidenced by their binding to sRAGE and esRAGE. These receptors include macrophage scavenger receptor types I and II (SR-A) [39], oligosaccharyl transferase-4 (OST-48 or AGE-R1) [40], Lectin-like oxidized LDL receptor-1 (LOX-1) [41], and protein (AGE-R3) [43]. The transmembrane receptor AGE-R1 or OST has an extracellular N-terminal domain and an intracellular C-terminal domain. It's known as a (translocon receptor) and is involved in the translocation of polypeptides across the membrane of eukaryotes [44], with AGE-R2 containing a tyrosine-

phosphorylated section anchored in the cell's plasma membrane and playing a role in intracellular signaling similar to the fibroblast growth factor receptor [42]. Patients with low levels of the soluble form of RAGE are more inclined to DM and cardiovascular illnesses, according to a research of 1201 participants done over 18 years [45].

The AGEs Role in the Pathogenesis of Diabetic Complications

Excessive AGEs production has been linked to microvascular and macrovascular problems in diabetic and non-diabetic patients. As a result, two approaches can be used to explain the pathophysiology of DM-related problems caused by high AGEs [46]. To begin with, AGEs can tangle with proteins and cause conformational changes, altering their functions and characteristics. Second, AGEs can trigger intracellular signals through receptor- or non-receptor-mediated pathways. Finally, these interactions result in an overproduction of inflammatory mediators such as cytokines and ROS [47-49]. AGEs have also been linked to atherosclerosis, since they reduced low-density lipoprotein clearance and increased the expression of a number of atherosclerosis-related molecules, including VEGF [50,49]. Furthermore, AGEs have the ability to interact with particular receptors (RAGEs), which are involved in the pathophysiology of DM complications. The development of the AGE-RAGE complex during hyperglycemia triggers a cascade of signals including TGF, NFkB, MAP kinase, and NADPH oxidases. As a result, E-selectin, vascular adhesion molecule-1, VEGF, and different pro-inflammatory cytokines such as IL-1 and IL-6 can be induced. TNF- α is highly induced by all of these signaling molecules. Vascular fibrosis, calcification, inflammation, prothrombotic effects, and vascular injury are all caused by them. These symptoms are comparable to those seen in diabetic nephropathy, neuropathy, retinopathy, and heart disease. Furthermore, when AGEs interact with macromolecules such as proteins, DNA, and the extracellular matrix (ECM), the structural conformation changes that ensue can have an impact on their biological function. As a result of the glycation process of DNA [51], the DNA-AGEs complex forms in diabetics, causing multi-neurological damage [52] and cancer [53]. The binding of AGEs to ECM changes the structure and biological behavior of ECM in DM. Meanwhile, crosslinking collagen I with AGEs alters tropocollagen's molecular structure and impairs normal tendon function [54,55].

DM-Related Cardiovascular Diseases

Cardiovascular disease is the leading cause of death among diabetics, particularly those with T2DM [56]. When diabetic individuals maintain their plasma glucose levels within acceptable ranges for up to 6 years, the risk of cardiovascular events decreases [57,58]. Patients with T2DM are four times more likely than non-diabetic patients to develop heart failure [59], and the mortality rate from diabetes cardiovascular complications is very high, reaching 75% of DM patients [60]. Nin *et al.* found that the occurrence of deadly and non-lethal cardiovascular events is associated with the blood level of AGEs during a 12-year period in a study of 339 diabetic patients [61]. Furthermore, Hassen and colleagues conducted a cohort study with a large number of T2DM patients and found that high levels of AGEs (e.g., CML, CEL, Pentosidine) are strongly associated with the prevalence of cardiovascular disease [62]. According to various studies conducted over the last seven years, a high AGEs/RAGEs ratio has a considerable impact on the development of aging-related disorders such as atherosclerosis [63], endothelial dysfunction [64], hyperthyroidism [65], and chronic renal failure [66].

AGEs in DM-induced Cardiovascular Disease: Mechanistic Issue

The accumulation of high AGEs in a chronic hyperglycemic state was linked to a higher incidence of cardiovascular diseases [67], most likely due to the initiation of oxidative stress [68], protein kinase (PKC) induction [69], chronic inflammatory reactions [70], mitochondrial dysfunction [71], and RAS activation [72] (Table 1).

Oxidative Stress

Oxidative stress is a condition in which the body's oxidation and antioxidation mechanisms are out of balance. Because of the stimulation of NADPH oxidase, xanthine oxidase (XO), and nitric oxide synthase (NOS), it may have a deleterious impact on several cellular functions [73]. The etiology of DM complications is complicated by oxidative stress [15]. Chronic hyperglycemia activates a NADPH-dependent oxidase, which catalyzes the production of ROS. It can lower anti-oxidant activity both enzymatically (superoxide dismutase) and non-enzymatically (ascorbic acid) [74]. As a result, oxidative stress causes inflammation, endothelial dysfunction, cardiomyocyte hypertrophy, and myocardial fibrosis, which leads to a decrease in left ventricular compliance, diastolic dysfunction, heart failure, arrhythmia, and/or sudden death.

Table 1. Summary of the clinical studies on the role of AGEs and RAGEs in diabetic cardiovascular complications

Author/year	Method	Participants	Result	Conclusion
Koska <i>et al.</i> (2018) [95]	Implication of AGEs and oxidative products in 1) a sub cohort Veterans Affairs Diabetes Trial (VADT), and 2) nested case-control subgroup from the ACCORD study.	VADT $n=445$ ACCORD $n=271$	In low MetOs, 107 participants show high CVD events, and highest incidence of CVD in the VADT study.	Individuals with long-term T2DM with low MetOs and high AGEs (glyoxal hydroimidazolones, or carboxymethyl lysine, or 3-deoxyglucosone hydroimidazolones) are more prone to CVD.
Saku <i>et al.</i> (2020) [97]	The AGEs-RAGE role in calcified AS pathophysiology was evaluated	$n=124$; 54 subjects with calcified AS; 70 control subjects without heart disease	Calcified AS remarkably had a higher level of AGEs, RAGEs, and RAGE expression than controlled subjects	AGEs and RAGE play a role in the pathophysiology of calcified AS, which act as a marker for calcified AS after aortic valve surgery.
McNair <i>et al.</i> (2016) [98]	Investigate the correlation between AGEs-RAGE and atherosclerosis in two groups of Caucasian patients	$n=95$; Divided into high cholesterol and normal cholesterol groups	sRAGE associates with HDL, whereas in hypercholesterolemia it has a passive role related to LDL, triglycerides, total cholesterol, and MDA.	A strong relationship observed between hypercholesterolemia and sRAGE, AGEs, MDA. Elevated sRAGEs increased HDL levels, conversely in the low level of sRAGEs.
Reichert <i>et al.</i> (2017) [99]	Longitudinal cohort study over 3 years, to determine the association of peripheral sRAGE with the CVD.	$n=1009$ individuals with CVD	MI 3.4%, stroke/TIA 2.4%, cardiac death 9.5%, death due to stroke 0.8%. The CVD recurrence was high in patients with sRAGE level >838.19 pg/ml.	Serum sRAGE impacted negatively CVD patients, and the research result disagrees with the hypothesis that presumes sRAGE has a protective role in CVD.
Manganelli <i>et al.</i> (2019) [100]	A cohort study in APS Caucasian patients to detect the correlation between high mobility (HMGB1)/sRAGE and the clinical manifestation in APS patients.	$n=90$; 60 subjects with APS: 25 with primary APS and 35 with APS+ SLE	In recurrent abortion individuals with APS, the serum of sRAGE elevated when compared with APS individuals without (control subject).	Both HMGB1 and sRAGEs were elevated in response to the increase in pro-inflammatory mediators and may have a role in autoimmune diseases. HMGB1 is useful in monitoring particular treatments such as antiaggregants.
Al Rifai <i>et al.</i> (2015) [101]	Community-based prospective cohort study. Evaluate the role of AGEs-RAGE in AF.	$n=1068$ participants	Patients with low sRAGE levels are more prone to develop AF.	There is an inverse relationship between sRAGE and inflammatory markers. No significant data support the role of sRAGE in the incidence of AF.

Role of AGEs in Atrial Stiffness

A 2018 study found a substantial link between AGEs and arterial wall stiffness, arrhythmias, systolic and diastolic dysfunction, congestive heart failure, coronary artery disorders, and the risk of in-stent restenosis [75]. The binding of the glycated forms of elastin and laminin with AGEs is responsible for all of these alterations. It has been linked to reducing nitric oxide (NO) generation through modifying cell-matrix interactions and weakening endothelial cell adhesion qualities [75]. Furthermore, the interaction of AGEs with cellular proteins that control intracellular Ca^{+2} levels (e.g., the sarcoendoplasmic reticulum Ca^{+2} -ATPase pump and the ryanoid receptor) alters Ca^{+2} concentration and disrupts cardiomyocyte contraction and relaxation, resulting in irregular diastolic tone [76].

Role of AGEs on Atherogenicity

AGEs can also crosslink with circulating biomolecules, changing their properties in recognizing

and processing receptors, such as the glycation of apolipoprotein B100, which is linked to foam cell generation and atherosclerosis due to a reduction in LDL receptor capacity and an increase in circulating LDL-c [77,75]. Glycation of fibroblasts also causes the fibrinolytic process to be disrupted [75].

Role of AGEs in Endothelial Dysfunction and Inflammation

The accumulation of AGEs in endothelial cells has been linked to a decline in endothelial function. It may play a role in the onset of cardiovascular diseases. In this context, AGEs suppress NOS expression and reduce NO production, resulting in the accumulation of asymmetric dimethyl arginine (ADMA). It will exacerbate the thrombotic tendency and accelerate the evolution of arteriosclerosis [75,78]. AGE-RAGE signaling also activates many intracellular pathways, resulting in the production of pro-inflammatory cytokines (e.g., IL-6, TNF- α , TGF- β), vascular adhesion molecules (VCAM-1, ICAM-1, ET-1), and ROS, all of which are linked to the establishment of

vascular inflammation [79]. Other receptors that AGEs can interact with include lipoxygenase-1 (LOX-1) and galactin-3 [80,81], and they can also boost LOX-1 expression, which is involved in atherosclerosis and cardiovascular diseases. The interaction of methylglyoxal hydroimidazolones 1 (MG-H1) with RAGEs upregulates ROS and intracellular adhesion molecules (ICAM) expression in human umbilical vein endothelial cells (HUVECs), according to Ishibashi *et al.* (2017). THP-1 (macrophage) adherence to HUVECs was also engaged in the inflammatory process and was thought to be an indication of atherosclerosis [82]. According to this findings, AGEs may play a role in cell damage by stimulating vascular endothelial growth factor (VEGF), plasminogen activator inhibitor (PAI-1), and thrombosis development [83].

Role of AGEs in Vascular Smooth Muscle Cells Function

The development of the AGEs-RAGEs complex changes the physiologic function of the vascular smooth muscles and contributes to atherosclerosis [84]. The biological cell cycle was disrupted by the buildup of AGEs in blood vessels and activation of the ERK/MAPK and Akt/mTOR pathways, which altered cellular proliferation by inducing death and autophagy [84]. Furthermore, RAGE expression enhances the formation of ROS, which activates NADPH oxidase and activates P38 MAPK [85], upregulates matrix metalloproteinase (MMP2/MMP9), and increases migratory capacity and lipocalin-2 expression [86]. Increased ROS expression promotes the transcription of receptor activator of NF κ -B ligand (RANKL), which leads to bone formation differentiation [87]. As a result, diabetic people are more likely to develop osteoporosis and have a higher risk of fractures [88,89].

Role of AGEs in Platelets Activation and Aggregation

Overexpression of AGEs and their interaction with RAGEs on the platelet membrane can affect platelet function, modifying platelet function through a cascade of intracellular events [90]. In this regard, AGEs promote platelet aggregation by modifying the phospholipid content of the cell membrane [91], increasing glycoprotein GPIIb [92], overexpressing ROS, and enhancing COX action and thromboxane A formation, which are the primary factors in microthrombus formation [93]. Maugeri and colleagues have revealed another platelet aggregation route involving the interaction of HMGB1 with neutrophil RAGE, which leads to activation of autophagolysosomes via the MAPK pathway [94].

DIABETIC NEPHROPATHY

Because of end-stage renal disease, diabetic nephropathy has a significant mortality and morbidity rate (102,103). Through auto-oxidation, glycosylation, and polyol pathways, both endogenous and exogenous AGEs contributed to affecting nephron biological processes and altering their structure [104]. Hyperglycemia promotes the synthesis of AGEs, which causes tubular endothelial cells to produce plasminogen activator inhibitor 1 and transglutaminase mRNA, allowing fibrosis-exacerbating macrophages to thrive [105]. Furthermore, activation of the AGEs receptor results in the production and release of pro-inflammatory cytokines and free radicals, which worsens nephropathic consequences by changing the glomerulus function [106]. In addition, lipid metabolism, RAS, systemic, and glomerular hypertension all had a role in the onset and progression of diabetic nephropathy [107]. Many investigations have shown that AGE levels in the blood are significantly linked to diabetic nephropathy, glomerulopathy, and mesangial cell proliferation [108,109]. After degradation by macrophages or extracellular proteolytic enzymes, AGEs are excreted through the renal pathway, yielding low molecular weight (LMW) AGEs (Table 2). However, high molecular weight AGEs may resist these enzymes by covering the enzymes' target site, resulting in the formation of LMW AGEs that are completely different from natural LMW AGEs [110-112].

DIABETIC NEUROPATHY

Diabetic neuropathy (DN) is a progressive disorder induced by chronic hyperglycemia that affects 30 to 50 percent of diabetic people [116]. It's possible that severe DM is linked to a loss of sensation in peripheral tissues, as well as an increased risk of infections like diabetic foot, which can lead to amputation [117]. Uncontrolled hyperglycemia contributes to the development of DN via several pathways, including the polyol route, the AGEs-RAGEs axis, the PKC pathway, and the hexosamine pathway [118,119]. Hyperglycemia activates the polyol pathway, in which glucose is converted to sorbitol by an aldose-reductase enzyme. This reaction depletes cellular NADPH, which lowers cytoplasmic antioxidants like glutathione (GSH) [120], inhibits the role of glyceraldehyde-3-phosphate dehydrogenase (GAPDH) in glycolysis pathways, and is linked to the activation of the hexosamine pathway through increased aggregation of GAPDH metabolites [119]. Furthermore, AGE accumulation and binding with RAGEs in nerves cause oxidative stress and the production of ROS, which leads to NF κ B-induced

apoptosis and neuronal injury [121]. Furthermore, AGE accumulation was linked to metabolic alterations in Schwann's cells, which led to cell death [122]. Diabetic neuropathy was strongly linked to PKC activation of PKC pathways by diacylglycerol after prolonged hyperglycemia [123]. Meanwhile,

aggregation of hexosamine pathway proteins has been associated to diabetic neuropathic problems, with dyslipidemia perhaps playing a role [124]. Al-Sofiani *et al.* reported a substantial link between sRAGE and high-frequency hearing loss in 2019 [125].

Table 2. Clinical and animal studies on the role of AGEs and RAGEs in diabetic nephropathy

Type of Study	Method	Participants	Result	Conclusion
Human study Normand <i>et al</i> (2018) [113]	Prospective, randomized, cross-over trial; evaluate the role of AGEs in renal hemodynamic variations. Renal perfusion and oxidative metabolism were assessed by PET [¹⁵ O]H ₂ O and [¹¹ C].	<i>n</i> = 10 healthy subjects	High-AGEs diet significantly increased renal perfusion, while low-AGEs diets do not show significant results. Oxygen consumption increased in high-AGEs diets.	Only high-AGEs contents are associated with renal hemodynamic changes with exacerbation of diabetic nephropathy.
Animal study Haraguchi <i>et al</i> (2020) [114]	Analyses of renal tubular lesions of early-stage diabetic kidney in (STZ)-induced DM animal model	<i>n</i> = 6 mice	STZ induces renal tubules apoptosis. AGEs accumulated in renal glomeruli and renal tubules, and RAGE expression increased compared with control kidneys.	The AGEs-RAGE axis is strongly associated with proximal renal tubules in the diabetic kidney.
Human study Perkins <i>et al.</i> (2020) [115].	Prospective case-control study	<i>n</i> = 85 subjects, 30 T1DM patients without albuminuria, and 22 patients with microalbuminuria and GFR decline, 33 subjects with microalbuminuria without GFR decline	Urinary excretion of pentosidine in MA subjects had a higher rate than NA subjects and higher in low GFR than normal GFR with or without MA. G-H1 and CMA increased and decreased respectively in MA with or without GFR decline.	The fraction excretion markedly higher in T1DM subjects with MA and decreased GFR compared with T1DM with or without MA. Elevated fraction excretion of AGEs may act as a clinical manifestation of depleted cation transport of the kidney.

DIABETIC RETINOPATHY

Diabetic retinopathy (DR) is a progressive condition that causes structural and functional loss of the retina and is the leading cause of blindness in adults and the elderly [126]. In the chronic condition, it is characterized by retinal vascular damage exhibited as blood-retina barrier breach and inducing the development of new vessels. Mild DR microaneurysms are the most common symptom; second, moderate DR with exudate, hemorrhage, and microvascular abnormalities; and third, severe DR microvascular damage with more than 20 hemorrhagic foci. Neovascularization was observed during the proliferative period, which results in neural and retinal vessel injury [128]. Retinal cells consume a lot of oxygen and are particularly vulnerable to oxidative stress and the generation of ROS, especially in hyperglycemic circumstances that change retinal structure and function. Inflammatory alterations, mitochondrial dysfunction, cell death by apoptosis, and gradual neurovascular injury of the retinal tissue

are all triggered by these processes [129]. The polyol pathway was activated during hyperglycemia, resulting in increased expression of the Aldoreductase enzyme, which is responsible for the synthesis and aggregation of sorbitol in the retina, causing edema and altering the blood-retinal-barrier function [130]. The expression of VEGF, which plays a key role in angiogenesis, increased permeability, and increases pro-inflammatory mediators, increased during the ischemia situation. All of these pathways have a role in DR pathogenesis [131]. Excessive production of AGEs, which are also involved in damaging retinal pericytes and bipolar cells [132], activates the hexosamine pathway, which negatively influences the connection between retinal pigmented endothelial cells, increases vascular permeability, and disrupts retinal vasculature [133,134]. Many studies have found that uncontrolled glycemic states are the primary cause of DR, and that managing hyperglycemia, blood pressure, and lipid profile has a good impact on reducing deterioration and the

occurrence of retinal complications [135]. By producing ROS through the stimulation of various processes such as polyol and hexosamine and PKC pathways, oxidative stress plays a significant role in the pathogenesis of DR [136]. The synthesis of sorbitol and the consumption of NADPH are facilitated in DM by activation of the polyol pathways. This has been linked to the activation of aldose reductase and sorbitol dehydrogenase [137], which causes NADPH levels to drop [138]. Obrosova *et al.* found low levels of NADPH and ATP in diabetic rats' lenses, while significant quantities of sorbitol and fructose were found [139]. New blood vessels proliferate and vascular permeability changes under hyperglycemic or hypoxic situations as a result of diacylglycerol (DAG), which is produced by the polyol pathway, stimulating the PKC pathway (PKC Ca⁺² and DAG dependent kinase) [140,141]. Because the rate of glycation rises in hyperglycemia, the synthesis of AGE precursors was abnormally high. Meanwhile, AGEs increased VEGF and pericyte apoptosis, and AGEs-RAGEs binding increases ROS production, depletes superoxide dismutase (SOD) and catalase, and alters the antioxidant activity of both GSH and ascorbic acid [142]. Furthermore, hyperglycemia can cause inflammatory reactions that raise cytokine levels as well as the creation of hydrogen peroxide, which helps NFκB develop. VEGF, COX2, MCP1 (monocyte chemotactic proteins), VCAM, and ICAM expression was considerably increased at this stage [131,143]. Meanwhile, COX2 increases the synthesis of PGs, stabilize HIF-1, and supports the expression of VEGF, NF-KB, and COX2; this pathway has been implicated in the formation of retinal aneurysms [144]. Furthermore, Tao *et al.* found a link between the hypoxic state of retinal micro-capillaries and the formation of AGEs, which predisposes to angiogenic processes [145]. Meanwhile, Kanda *et al.* discovered AGE accumulation at the optic peripheral nerve [146-148]. AGEs were found to play a significant role in the pathogenesis of several ocular illnesses in the cornea [149-153], lens [154-157], vitreous humor [158,159], and optic nerve [160,161], according to several investigations.

BONE METABOLISM

Diabetics are more likely to fracture bones because persistent hyperglycemia changes bone metabolism, composition, and inhibits fracture healing [162]. Due to increased levels of glycated collagen and AGE aggregation in the bone, many variables contribute to altered bone metabolism and bone dysfunction (e.g., oxidative stress, depleted IGF-1, an elevated level of Sclerotin, and decreased bone density) [162,163]. The

activation and potentiation of inflammatory and oxidative stress pathways by the AGE-RAGE axis played a key role in the impairment of bone metabolism [164]. Furthermore, the AGEs-RAGEs axis and RAGEs ligands disrupt bone remodeling [165], as well as cytokines (TGF-β and IGF-1) during osteoblast development [166]. AGEs (Pentosidine) have been discovered in the bone cells of diabetic patients [167]. Both osteoclasts and osteoblasts are affected by AGEs, with osteoclastogenesis induced by overexpression of RANKL mRNA and osteoblast downregulation impacting the mineralization process [168-170]. Furthermore, depending on the concentration of human fetal osteoblastic cells (hFOB), AGEs have a protective and destructive effect on hFOB cells. At low concentrations, it preserves hFOB cells by inducing osteoblastic cells to function and dropping osteoclastic function cells out, whereas at high concentrations, it has the opposite effect [171]. Glycated collagen has a lower ability to attach to osteoblasts via discoidin domain receptors (DDR2) and integrin receptors [172], resulting in lower lysyl oxidase synthesis. AGEs enhance collagen synthesis, while a decrease in collagen leads to rapid breakdown [173]. AGEs-modified proteins, on the other hand, interact with RAGEs to limit collagen synthesis in fibroblasts; moreover, increased pro-inflammatory cytokines affect collagenase production [174]. In diabetic individuals, resistance to glycated collagen degradation causes a decrease in the amount of C-telopeptides of type 1 collagen (CTX-1), as well as depletion of osteoblast, and a decrease in collagen synthesis causes a decrease in the level of N-terminal propeptide of type I collagen (P1NP) [175]. Table 3 summarizes various human studies on the effects of diabetes on bone metabolism.

Conclusion

AGEs are heterogeneous chemical compounds that are formed when sugar reacts with macromolecules, either enzymatically or non-enzymatically. RAGE develops as a result of intrinsic cellular signaling induced by AGEs. The formation of AGE/RAGE and diabetes complications share a number of risk factors. The inhibition of several pathogenic alterations in the setting of DM could be due to an AGE-RAGE interaction. sRAGE levels can be raised as a result of excessive RAGE production or degradation, and it doesn't always reflect AGE-RAGE signaling. Targeting the AGEs-sRAGE axis (Table 4) could be a good strategy to reduce the consequences of the AGE-RAGE signaling pathway in many tissues and organs damaged by long-term hyperglycemia, however it can't be used in clinical practice due of the limitations.

Table 3. Clinical studies on the role of AGEs and RAGEs in DM-induced disturbance of bone metabolism

Author/year	Method	Number of participants	Result	Conclusion
Rabelo <i>et al.</i> (2018) [176]	Analysis of cortical fractal dimension and lacunarity, and collagen crosslinks content in cortical bone.	35 postmenopausal women femoral neck sample; 17 with fracture; 18 with OA and carcinoma.	Increase in the pentosidine femoral neck of osteoporotic fractures independent of age.	Bone quality and risk of hip fracture are associated with the level of Pentosidine. Increased pentosidine worsen the quality of bone and the incidence of hip fracture increased.
Tamaki <i>et al.</i> (2018) [177].	A prospective cohort study to evaluate the utility of serum pentosidine and esRAGE levels as predictors of fragility fractures.	1,285 Japanese men aged ≥ 65 years	The crude fragility fracture HRs (95% CI) PEN 1.56 (1.05–2.31) esRAGE 0.79 (0.54–1.15) esRAGE/PEN 0.65(0.44–0.95).	The high level of sRAGE/pentosidine was associated with a reduced risk of fractures.
Vaculik <i>et al.</i> (2016) [178].	A cohort study to measure serum & bone pentosidine levels in femoral neck fracture and advanced hip osteoarthritis using HPLC.	111 patients undergoing hip surgery; 70 with a femoral neck fracture and 41 with advanced hip osteoarthritis.	Serum and bone pentosidine levels were elevated in both fractured and arthritis patients.	There is a significant relationship between pentosidine level and hip fracture and osteoarthritis. Pentosidine can be a biomarker for assessing the quality of bone.
Furst <i>et al.</i> (2016) [88]	A cross-sectional study to determine the impact of AGEs on bone materials strength in T2DM patients.	35 postmenopausal women; 16 with T2DM and 19 controls.	AGEs level was highly correlated with decreased bone material strength and lower bone formation marker.	T2DM alters bone materials strength.
Farley <i>et al.</i> 2016 [179]	A cross-sectional, case-control study to determine whether bone matrix from fracturing and non-fracturing T1DM contained more AGEs than bone from healthy patients.	15 subjects; 5 fractured with T1DM; 5 T1DM without fracture; 5 healthy subjects	The fractured with T1DM had a high level of AGEs (Pentosidine) than the other groups.	Bone structure and density in T1DM are affected by a high level of AGEs (Pentosidine).

Acknowledgement

The authors thank College of Pharmacy, University of Sulaimani for supporting the project.

Conflicting interests

Nothing declared.

Data sharing statement

N/A

Table 4. Studies on pharmacological targeting of the AGEs-RAGEs axis to treat diabetes mellitus

Medication	Targets	Type of Study	Study Outcomes
Spironolactone [115]	AGEs-RAGEs axis inhibitors	An animal study using male Sprague-Dawley rats	Protect against diabetic-nephropathy by hindering the AGEs-RAGEs complex and increase SIRT-3 expression.
Catalpol (Iridous glucoside) [116]	Anti-inflammatory Anti-apoptotic Anti-oxidant	Review of 100 publications	Reduces nephropathy, cardiopathy, and neuropathy.
Resveratrol [117]	Anti-inflammatory Antioxidant	Review of 21 clinical trials	Attenuates production of IL-1 β , iL-6, TNF- α , and inducible nitric oxide synthase, and COX2
Glucopyranoside [118]	AGEs formation inhibitor	Animal study, Zebrafish	Potent antiglycative
Irbesartan [119]	Angiotensin II receptor antagonist	An animal study using male rats	Decrease cardiopathy through decreasing AGEs and expression of RAGEs.
Fluorofenidone [120]	Inhibitor of p38 MAPK, TNF- α , and TGF- β .	Animal Study using four weeks old mice	Alleviates nephropathy reducing ROS and modulating the mitochondrial function.
Hesperetin [119]	Inhibits AGEs formation with anti-inflammatory activity	An animal study using male rats	Nephroprotection via inhibiting AGEs formation, and decrease RAGEs expression in endothelial cells.
Interleukin-10 [119]	Anti-inflammatory Immune-regulatory	Animal study using male rats	Preserves the Schwann's cells by inhibiting the phosphorylation of NF- κ B.
SGLT-2 inhibitor, phlorizin analogs [121]	Na ⁺ -glucose cotransporter 2 inhibitor	A meta-Analysis of five large clinical trials	Cardio and renal protection via inhibition of the AGE-RAGE axis.
Zafirlukast [122]	Interleukin receptor antagonist	Human primary chondrocytes	Attenuates the development and progression of OA by decreasing AGEs
FPS-ZM1 and Valsartan [123]	Selective RAGE inhibitor and ARB	An animal study using male rats	Synergistic effect to prevent worsening of diabetic nephropathy.
Pyridoxamine (B6) [124]	Glycation process inhibitor	An animal study using male rats	Blocks RAGE-NF- κ B/ERK pathway and attenuates diabetic neuropathic pain.
Aminoguanidine [125]	AGEs and advanced oxidative protein product inhibitor	An animal study using male rats	Preserves against cardiac fibrosis through interrupting the AGEs-RAGE axis.
Glycine [180]	Antioxidant	An animal study using rats	Restrains production of AGEs and inhibits the RAGE.
Metformin [181]	Antihyperglycemic agent	Human study	Blocks AGEs formation; decreases inflammation, and elevates the esRAGE.
Dulaglutide GLP-1 analog [182]	Antidiabetic agent	In vitro study using hSW1353 cell line.	Regulates AGE-induced inflammation and damage in chondrocytes.
Vitamin D [183]	Supplement	A clinical trial	Attenuates the onset of cardiovascular complications after 3 months.

REFERENCES

- Poulsen MW, Hedegaard RV, Andersen JM, Courten BD, Bügel S, Nielsen J, et al. Advanced glycation endproducts in food and their effects on health. *Food Chem Toxicol.* 2013;60(10):10-37. doi: 10.1016/j.fct.2013.06.052.
- Vos T, Abajobir AA, Abbafati C, Abbas KM, Abate KH, Abd-Allah F, et al. GBD 2016 Disease and Injury Incidence and Prevalence Collaborators. Global, regional, and national incidence, prevalence, and years lived with disability for 328 diseases and injuries for 195 countries, 1990-2016: a systematic analysis for the Global Burden of Disease Study 2016. *Lancet.* 2017;390(10100):1211-1259. doi: 10.1016/S0140-6736(16)31678-6.
- Cho NH, Shaw JE, Karuranga S, Huang Y, da Rocha Fernandes JD, Ohlrogge AW, et al. IDF Diabetes Atlas: Global estimates of diabetes prevalence for 2017 and projections for 2045. *Diabetes Res Clin Pract.* 2018;138:271-281. doi: 10.1016/j.diabres.2018.02.023.
- Basu S, Yudkin JS, Kehlenbrink S, Davies JI, Wild SH, Lipska KJ, et al. Estimation of global insulin use for type 2 diabetes, 2018-30: A microsimulation analysis. *Lancet Diabetes*

- Endocrinol.* 2019;7(1):25-33. doi: 10.1016/S2213-8587(18)30303-6.
5. Ogurtsova K, da Rocha Fernandes JD, Huang Y, Linnenkamp U, Guariguata L, Cho NH, et al. IDF Diabetes Atlas: Global estimates for the prevalence of diabetes for 2015 and 2040. *Diabetes Res Clin Pract.* 2017;128:40-50. doi: 10.1016/j.diabres.2017.03.024.
 6. Paneni F, Luscher TF. Cardiovascular protection in the treatment of type 2 diabetes: a review of clinical trial results across drug classes. *Am J Cardiol.* 2017;120(1S):S17-S27. doi: 10.1016/j.amjmed.2017.04.008.
 7. Sardu C, De Lucia C, Wallner M, Santulli G. Diabetes mellitus, and its cardiovascular complications: new insights into an old disease. *J Diabetes Res.* 2019;2019:1905194. doi: 10.1155/2019/1905194.
 8. Vlassara H, Palace MR. Diabetes and advanced glycation end products. *J Intern Med.* 2002;251:87-101. doi: 10.1046/j.1365-2796.2002.00932.x.
 9. Bate KL, Jerums G. Preventing complications of diabetes. *Med J Aust.* 2003;179:498-503. doi: 10.5694/j.1326-5377.2003.tb05655.x.
 10. Yang J, Deng S, Yin J, Yu J, Chu G, Cui H, et al. Preparation of 1-amino-1- deoxyfructose derivatives by stepwise increase of temperature in aqueous medium and their flavor formation compared with Maillard reaction products. *Food Bioprocess Technol.* 2018; 11(3):694-704. doi: 10.1007/s11947-017-2039-4.
 11. Oliveira FCD, Coimbra JSJR, Oliveira EBD, Zuñiga, ADG, Rojas EEG. Food protein-polysaccharide conjugates obtained via the Maillard reaction: A review. *Crit Rev Food Sci Nutr.* 2016;56(7):1108-1125. doi: 10.1080/10408398.2012.755669.
 12. Sharma C, Kaur A, Thind SS, Singh B, Raina S. Advanced glycation endproducts (AGEs): An emerging concern for processed food industries. *J Food Sci Technol.* 2015;52(12):7561-7576. doi: 10.1007/s13197-015-1851-y.
 13. Martins SIFS, Jongen WMF, Boekel MAJSV. A review of Maillard reaction in food and implications to kinetic modelling. *Trends Food Sci Technol.* 2000;11(9-10):364-373. doi: 10.1016/S0924-2244(01)00022-X.
 14. Fournet M, Bonté F, Desmoulière A. Glycation damage: A possible hub for major pathophysiological disorders and aging. *Aging Dis.* 2018;9(5):880-900. doi:10.14336/AD.2017.1121.
 15. Faria A, Persaud SJ. Cardiac oxidative stress in diabetes: mechanisms and therapeutic potential. *Pharmacol Ther.* 2017;172:50-62. doi: 10.1016/j.pharmthera.2016.11.013.
 16. Davis KE, Prasad C, Vijayagopal P, Juma S, Imrhan V. Advanced glycation end products, inflammation, and chronic metabolic diseases: Links in a chain? *Crit Rev Food Technol Nutr.* 2016;56(6):989-998. doi: 10.1080/10408398.2012.744738.
 17. Tessier FJ, Niquet-Léridon C, Jacolot P, Jouquand C, Genin M, Schmidt AM, et al. Quantitative assessment of organ distribution of dietary protein-bound ¹³C-labeled N^ε-carboxymethyllysine after a chronic oral exposure in mice. *Mol Nutr Food Res.* 2016;60(11):2446-2456. doi: 10.1002/mnfr.201600140.
 18. Van Nguyen C. Toxicity of the AGEs generated from the Maillard reaction: on the relationship of food-AGEs and biological-AGEs. *Mol Nutr Food Res.* 2006;50:1140-1149. doi: 10.1002/mnfr.200600144.
 19. Thorpe SR, Baynes JW. Maillard reaction products in tissue proteins: new products and new perspectives. *Amino Acids.* 2003;25:275-281. doi: 10.1007/s00726-003-0017-9.
 20. Koschinsky T, He CJ, Mitsuhashi T, Bucala R, Liu C, Buenting C, et al. Orally absorbed reactive glycation products (glycotoxins): An environmental risk factor in diabetic nephropathy. *Proc Natl Acad Sci.* 1997;94:6474-6479. doi: 10.1073/pnas.94.12.6474.
 21. Uribarri J, Woodruff S, Goodman S, Cai W, Chen X, Pyzik R, et al. 2010. Advanced glycation end products in foods and a practical guide to their reduction in the diet. *J Am Dietetic Assoc.* 2010;110:911-916. doi: 10.1016/j.jada.2010.03.018.
 22. Byun K, Yoo YC, Son M, Lee J, Jeong GB, Park YM, et al. Advanced glycation end products produced systemically and by macrophages: a common contributor to inflammation and degenerative diseases. *Pharmacol Ther.* 2017;177:44-55. doi: 10.1016/j.pharmthera.2017.02.030.
 23. Reynaert NL, Gopal P, Rutten EPA, Wouters EFM, Schalkwijk CG. Advanced glycation end products and their receptor in age-related, non-communicable chronic inflammatory diseases; Overview of clinical evidence and potential contributions to disease. *Int J Biochem Cell Biol.* 2016;81(Pt B):403-418. doi: 10.1016/j.biocel.2016.06.016.
 24. Xue J, Rai V, Singer D, Chabierski S, Xie J, Reverdatto S, et al. Advanced glycation end product recognition by the receptor for AGEs. *Structure.* 2011;19(5):722-732. doi: 10.1016/j.str.2011.02.013.
 25. Schmoch T, Uhle F, Siegler BH, Fleming T, Morgenstern J, Nawroth PP, et al. The glyoxalase system and methylglyoxal-derived carbonyl stress in sepsis: Glycotoxic aspects of sepsis pathophysiology. *Int J Molecular Sci.* 2017;18(3):657. doi: 10.3390/ijms18030657.
 26. Lopez-Moreno J, Quintana-Navarro GM, Camargo A, Jimenez-Lucena R, Delgado-Lista J, Marin C, et al. Dietary fat quantity and quality modifies advanced glycation end products metabolism in patients with metabolic syndrome. *Mol Nutr Food Res.* 2017;61(8):1601029. doi: 10.1002/mnfr.201601029.
 27. Lin JA, Wu CH, Lu CC, Hsia SM, Yen GC. Glycative stress from advanced glycation end products (AGEs) and dicarbonyls: An emerging biological factor in cancer onset and progression. *Mol Nutr Food Res.* 2016;60(8):1850-1864. doi: 10.1002/mnfr.201500759.
 28. Delgado-Andrade C, Fogliano V. Dietary advanced glycosylation end-products (dAGEs) and melanoidins formed through the Maillard reaction: Physiological consequences of their intake. *Ann Rev Food Sci Technol.* 2018;9(1):271-279. doi: 10.1146/annurev-food-030117-012441.
 29. Eisermann DJ, Wenzel U, Fitzenberger E. Inhibition of chaperone-mediated autophagy prevents glucotoxicity in the *Caenorhabditis elegans mev-1* mutant by activation of the proteasome. *Biochem Biophys Res Commun.* 2017;484:171-175. doi: 10.1016/j.bbrc.2017.01.043.
 30. Xue J, Ray R, Singer D, Böhme D, Burz DS, Rai V, et al. The receptor for advanced glycation end products (RAGE) specifically recognizes methylglyoxal-derived AGEs. *Biochemistry.* 2014;53(20):3327-3335. doi: 10.1021/bi500046t.

31. Fritz G. RAGE: A single receptor fits multiple ligands. *Trends Biochem. Sci.* 2011;36:625-632. doi: 10.1016/j.tibs.2011.08.008.
32. Schmidt AM. Soluble RAGEs-prospects for treating and tracking the metabolic and inflammatory disease. *Vasc Pharmacol.* 2015;72:1-8. doi: 10.1016/j.vph.2015.06.011.
33. Kamo T, Tasaka S, Tokuda Y, Suzuki S, Asakura T, Yagi K, et al. Levels of soluble receptor for advanced glycation end products in bronchoalveolar lavage fluid in patients with various inflammatory lung diseases. *Clin Med Insights Circ Respir Pulmonary Med.* 2016; 9(Suppl. 1):147-154. doi: 10.4137/CCRPM.S23326.
34. Park IH, Yeon SI, Youn JH, Choi JE, Sasaki N, et al. Expression of a novel secreted splice variant of the receptor for advanced glycation end products (RAGE) in human brain astrocytes and peripheral blood mononuclear cells. *Mol Immunol.* 2004;40:1203-1211. doi: 10.1016/j.molimm.2003.11.027.
35. Zhang L, Bukulin M, Kojro E, Roth A, Metz VV, Fahrenholz F, et al. Receptor for advanced glycation end products is subjected to protein ectodomain shedding by metalloproteinases. *J Biol Chem.* 2008;283:35507-35516. doi: 10.1074/jbc.M806948200.
36. Metz VV, Kojro E, Rat D, Postina R. Induction of RAGE shedding by activation of G protein-coupled receptors. *PLoS ONE.* 2012;7:e41823. doi: 10.1371/journal.pone.0041823.
37. Yonekura H, Yamamoto Y, Sakurai S, Watanabe T, Yamamoto H. Roles of the receptor for advanced glycation endproducts in diabetes-induced vascular injury. *J Pharmacol Sci.* 2005; 97:305-311. doi: 10.1254/jphs.cpj04005x.
38. Srikrishna G, Huttunen HJ, Johansson L, Weigle B, Yamaguchi Y, et al. N-Glycans on the receptor for advanced glycation end products influence amphoterin binding and neurite outgrowth. *J Neurochem.* 2002;80:998-1008. doi: 10.1046/j.0022-3042.2002.00796.x.
39. Yang Z, Makita Z, Hori Y, Brunelle S, Cerami A, Sehajpal P, et al. Two novel rat liver membrane proteins that bind advanced glycosylation end products: relationship to macrophage receptor for glucose-modified proteins. *J Exp Med.* 1991;174(3):515-524. doi: 10.1084/jem.174.3.515.
40. Kelleher DJ, Kreibich G, Gilmore R. Oligosaccharyl transferase activity is associated with a protein complex composed of ribophorins I and II and a 48 kd protein. *Cell.* 1992;69:55-65. doi: 10.1016/0092-8674(92)90118-v.
41. Franke S, Dawczynski J, Strobel J, Niwa T, Stahl P, Stein G. Increased levels of advanced glycation end products in human cataractous lenses. *J Cataract Refract Surg.* 2003;29:998-1004. doi: 10.1016/s0886-3350(02)01841-2.
42. Horiuchi S, Sakamoto Y, Sakai M. Scavenger receptors for oxidized and glycated proteins. *Amino Acids.* 2003;25:283-292. doi: 10.1007/s00726-003-0029-5.
43. Vlassara H, Li YM, Imani F, Wojciechowicz D, Yang Z, Liu FT, et al. Identification of galectin-3 as a high-affinity binding protein for advanced glycation end products (AGE): a new member of the AGE-receptor complex. *Mol Med.* 1995;1(6):634-646. PMID: 8529130
44. Krieg UC, Johnson AE, Walter P. Protein translocation across the endoplasmic reticulum membrane: identification by photo cross-linking of a 39-kD integral membrane glycoprotein as part of a putative translocation tunnel. *J Cell Biol.* 1989;109:2033-2043. doi: 10.1083/jcb.109.5.2033.
45. Selvin E, Halushka MK, Rawlings AM, Hoogeveen RC, Ballantyne CM, Coresh J, et al. sRAGE and risk of diabetes, cardiovascular disease, and death. *Diabetes.* 2013;62(6):2116-2121. doi: 10.2337/db12-1528.
46. Vlassara H, Uribarri J. Advanced glycation end products (AGE) and diabetes: cause, effect, or both? *Curr Diab Rep.* 2014;14:453. doi: 10.1007/s11892-013-0453-1.
47. Hofmann MA, Drury S, Fu C, Qu W, Taguchi A, Lu Y, et al. RAGE mediates a novel proinflammatory axis: the cell surface receptor for S100/calgranulin polypeptides. *Cell.* 1999; 97:889-901. doi: 10.1016/s0092-8674(00)80801-6.
48. Hori O, Brett J, Slattery T, Cao R, Zhang J, Chen JX, et al. The receptor for advanced glycation end products (RAGE) is a cellular binding site for amphoterin. Mediation of neurite outgrowth and co-expression of rage and amphoterin in the developing nervous system. *J Biol Chem.* 1995;270(43):25752-25761. doi: 10.1074/jbc.270.43.25752.
49. Yamagishi S. Role of advanced glycation end products (AGEs) and receptor for AGEs (RAGE) in vascular damage in diabetes. *Exp Gerontol.* 2011;46(4):217-224. doi: 10.1016/j.exger.2010.11.007.
50. Bucala R, Mitchell R, Arnold K, Innerarity T, Vlassara H, Cerami A. Identification of the major site of apolipoprotein B modification by advanced glycosylation end products blocking uptake by the low-density lipoprotein receptor. *J Biol Chem.* 1995;270(18):10828-10832. doi: 10.1074/jbc.270.18.10828.
51. Jaramillo R. DNA advanced glycation end products (DNA-AGEs) are elevated in urine and tissue in an animal model of type 2 diabetes. *Chem Res Toxicol.* 2017;30:689-698. doi: 10.1021/acs.chemrestox.6b00414.
52. Akhter F, Akhter A, Ahmad S. Toxicity of protein and DNAAGEs in neurodegenerative diseases (NDDs) with decisive approaches to stop the deadly consequences. In *Perspectives in Environmental Toxicology*. Environmental Science and Engineering, Kesari K, ed. (Springer), pp. 99–134, 2017. doi: 10.1007/978-3-319-46248-6_5.
53. Jandial R, Neman J, Lim PP, Tamae D, Kowolik CM, Wuenschell GE, et al. Inhibition of GLO1 in glioblastoma multiforme increases DNA-AGEs, stimulates RAGE expression, and inhibits brain tumor growth in orthotopic mouse models. *Int J Mol Sci.* 2018;19(2):406. doi: 10.3390/ijms19020406.
54. Hudson DM, Archer M, King KB, Eyre DR. Glycation of type I collagen selectively targets the same helical domain lysine sites as lysyl oxidase-mediated cross-linking. *J Biol Chem.* 2018;293(40):15620-15627. doi: 10.1074/jbc.RA118.004829.
55. Gautieri A, Passini FS, Silván U, Guizar-Sicairos M, Carimati G, Volpi P, et al. Advanced glycation end-products: Mechanics of aged collagen from molecule to tissue. *Matrix Biol.* 2017;59:95-108. doi: 10.1016/j.matbio.2016.09.001.
56. Sarwar N, Gao P, Seshasai SR, Gobin R, Kaptoge S, Di Angelantonio E, et al. Diabetes mellitus, fasting blood glucose concentration, and risk of vascular disease: a collaborative meta-analysis of 102 prospective studies. *Lancet.* 2010;375(9733):2215-22. doi: 10.1016/S0140-6736(10)60484-9.
57. Patel A, MacMahon S, Chalmers J, Neal B, Billot L, Woodward M, et al. Intensive blood glucose control and vascular

- outcomes in patients with type 2 diabetes. *N Engl J Med.* 2008;358(24):2560-2572. doi: 10.1056/NEJMoa0802987.
58. Duckworth W, Abraira C, Moritz T, Reda D, Emanuele N, Reaven PD, et al. Glucose control and vascular complications in veterans with type 2 diabetes. *N Engl J Med.* 2009;360(2):129-139. doi: 10.1056/NEJMoa0808431.
 59. Hangaard MH, Rossing P, Jensen JS, Jensen MT. Heart failure often accompanies diabetes mellitus. *Ugeskr Laeger.* 2018;180(20A). PMID: 30274586
 60. Umamahesh K, Vigneswari A, Surya Thejaswi G, Satyavani K, Viswanathan V. Incidence of cardiovascular diseases and associated risk factors among subjects with type 2 diabetes - an 11-year follow up study. *Indian Heart J.* 2014;66(1):5-10. doi: 10.1016/j.ihj.2013.12.009.
 61. Nin JW, Jorsal A, Ferreira I, Schalkwijk CG, Prins MH, Parving HH, et al. Higher plasma levels of advanced glycation end products are associated with incident cardiovascular disease and all-cause mortality in type 1 diabetes: a 12-year follow-up study. *Diabetes Care.* 2011;34(2):442-447. doi: 10.2337/dc10-1087.
 62. Hanssen NM, Beulens JW, van Dieren S, Scheijen JL, van der A DL, Spijkerman AM, et al. Plasma advanced glycation end products are associated with incident cardiovascular events in individuals with type 2 diabetes: a case-cohort study with a median follow-up of 10 years (EPIC-NL). *Diabetes.* 2015;64(1):257-265. doi: 10.2337/db13-1864.
 63. Tahara N, Yamagishi S, Tahara A, Ishibashi M, Hayabuchi N, Takeuchi M, et al. Adiponectin is inversely associated with ratio of serum levels of AGEs to sRAGE and vascular inflammation. *Int J Cardiol.* 2012;158(3):461-462. doi: 10.1016/j.ijcard.2012.05.015.
 64. Kajikawa M, Nakashima A, Fujimura N, Maruhashi T, Iwamoto Y, Iwamoto A, et al. Ratio of serum levels of AGEs to soluble form of RAGE is a predictor of endothelial function. *Diabetes Care.* 2015;38(1):119-125. doi: 10.2337/dc14-1435.
 65. Caspar-Bell G, Dhar I, Prasad K. Advanced glycation end products (AGEs) and its receptors in the pathogenesis of hyperthyroidism. *Mol Cell Biochem.* 2016;414(1-2):171-178. doi: 10.1007/s11010-016-2669-2.
 66. Prasad K, Dhar I, Zhou Q, Elmoselhi H, Shoker M, Shoker A. AGEs/sRAGE, a novel risk factor in the pathogenesis of end-stage renal disease. *Mol Cell Biochem.* 2016;423(1-2):105-114. doi: 10.1007/s11010-016-2829-4.
 67. Jia G, Whaley-Connell A, Sowers JR. Diabetic cardiomyopathy: a hyperglycemia- and insulin-resistance-induced heart disease. *Diabetologia.* 2018;61(1):21-28. doi: 10.1007/s00125-017-4390-4.
 68. Jha JC, Ho F, Dan C, Jandeleit-Dahm K. A causal link between oxidative stress and inflammation in cardiovascular and renal complications of diabetes. *Clin Sci.* 2018;132(16):1811-1836. PMID: 30166499
 69. Idris I, Gray S, Donnelly R. Protein kinase C activation: isozyme-specific effects on metabolism and cardiovascular complications in diabetes. *Diabetologia.* 2001;44(6):659-673. doi: 10.1007/s001250051675.
 70. Pedicino D, Liuzzo G, Trotta F, Giglio AF, Giubilato S, Martini F, et al. Adaptive immunity, inflammation, and cardiovascular complications in type 1 and type 2 diabetes mellitus. *J Diabetes Res.* 2013;2013:184258. doi: 10.1155/2013/184258.
 71. Vernochet C, Damilano F, Mourier A, Bezy O, Mori MA, Smyth G, et al. Adipose tissue mitochondrial dysfunction triggers a lipodystrophic syndrome with insulin resistance, hepatosteatosis, and cardiovascular complications. *FASEB J.* 2014;28(10):4408-19. doi: 10.1096/fj.14-253971.
 72. Hayashi T, Takai S, Yamashita C. Impact of the renin-angiotensin-aldosterone system on cardiovascular and renal complications in diabetes mellitus. *Curr Vasc Pharmacol.* 2010;8(2):189-197. doi: 10.2174/157016110790886947.
 73. Hu XF, Wang L, Xiang G, Lei W, Feng YF. Angiogenesis impairment by the NADPH oxidase- triggered oxidative stress at the bone-implant interface: critical mechanisms and therapeutic targets for implant failure under hyperglycemic conditions in diabetes. *Acta Biomater.* 2018;73:470-487. doi: 10.1016/j.actbio.2018.04.008.
 74. Suriyaprom K, Kaewprasert S, Putpadungwipon P, Namjuntra P, Klongthay S. Association of antioxidant status and inflammatory markers with metabolic syndrome in Thais. *J Health Population Nutr.* 2019;38(1):1. doi: 10.1186/s41043-018-0158-9.
 75. Fishman SL, Sonmez H, Basman C, Singh V, Poretzky L. The role of advanced glycation end-products in the development of coronary artery disease in patients with and without diabetes mellitus: a review. *Mol Med.* 2018;24:59. doi: 10.1186/s10020-018-0060-3.
 76. Bidasee KR, Zhang Y, Shao CH, Wang M, Patel KP, Dincer UD, et al. Diabetes increases formation of advanced glycation end products on Sarco(endo)plasmic reticulum Ca²⁺-ATPase. *Diabetes.* 2004;53(2):463-473. doi: 10.2337/diabetes.53.2.463.
 77. Yamagishi SI, Matsui T. Role of hyperglycemia-induced advanced glycation end product (AGE) accumulation in atherosclerosis. *Ann Vasc Dis.* 2018;11(2018):253-258. doi: 10.3400/avd.ra.18-00070.
 78. Ando R, Ueda S, Yamagishi S, Miyazaki H, Kaida Y, Kaifu K, et al. Involvement of advanced glycation end product-induced asymmetric dimethyl arginine generation in endothelial dysfunction. *Diab Vasc Dis Res.* 2013;10:436-441. doi: 10.1177/1479164113486662.
 79. Yamagishi SI, Matsui T. Role of hyperglycemia-induced advanced glycation end product (AGE) accumulation in atherosclerosis. *Ann Vasc Dis.* 2018;11:253-258. doi: 10.3400/avd.ra.18-00070.
 80. Lubrano V, Balzan S. Roles of LOX-1 in microvascular dysfunction. *Microvasc Res.* 2016;105:132-140. doi: 10.1016/j.mvr.2016.02.006.
 81. Yang P, Feng J, Peng Q, Liu X, Fan Z. Advanced glycation end products: Potential mechanism and therapeutic target in cardiovascular complications under diabetes. *Oxid Med Cell Longev.* 2019;2019:9570616. doi: 10.1155/2019/9570616.
 82. Ishibashi Y, Matsui T, Nakamura N, Sotokawauchi A, Higashimoto Y, Yamagishi SI. Methylglyoxal-derived hydroimidazolones-1 evokes inflammatory reactions in endothelial cells via an interaction with receptor for advanced glycation end products. *Diab Vasc Dis Res.* 2017;14:450-453. doi: 10.1177/1479164117715855.
 83. Li Z, Zhong Q, Yang T, Xie X, Chen M. The role of profilin-1 in endothelial cell injury induced by advanced glycation end products (AGEs). *Cardiovasc Diabetol.* 2013;12:141. doi: 10.1186/1475-2840-12-141.

84. Ma M, Guo X, Chang Y, Li C, Meng X, Li S, et al. Advanced glycation end products promote proliferation and suppress autophagy via reduction of Cathepsin D in rat vascular smooth muscle cells. *Mol Cell Biochem.* 2015;403:73-83. doi: 10.1007/s11010-015-2338-x.
85. Hu P, Lai D, Lu P, Gao J, He H. ERK and Akt signaling pathways are involved in advanced glycation end product-induced autophagy in rat vascular smooth muscle cells. *Int J Mol Med.* 2012;29:613-618. doi: 10.3892/ijmm.2012.891.
86. Dhar S, Sun Z, Meiningner GA, Hill MA. Nonenzymatic glycation interferes with fibronectin-integrin interactions in vascular smooth muscle cells. *Microcirculation.* 2017;24(3):12347. doi: 10.1111/micc.12347.
87. Menini S, Iacobini C, Ricci C, Fantauzzi CB, Salvi L, Pesce CM, et al. The galectin-3/RAGE dyad modulates vascular osteogenesis in atherosclerosis. *Cardiovasc Res.* 2013;100:472-480. doi: 10.1093/cvr/cvt206.
88. Furst JR, Bandeira LC, Fan WW, Agarwal S, Nishiyama KK, McMahon DJ, et al. Advanced glycation end products and bone material strength in type 2 diabetes. *J Clin Endocrinol Metab.* 2016;101(6):2502-2510. doi: 10.1210/jc.2016-1437.
89. Cheng YZ, Yang SL, Wang JY, Ye M, Zhuo XY, Wang LT, et al. Irbesartan attenuates advanced glycation end products-mediated damage in diabetes-associated osteoporosis through the AGEs/RAGE pathway. *Life Sci.* 2018;205:184-192. doi: 10.1016/j.lfs.2018.04.042.
90. Xie J, Méndez JD, Méndez-Valenzuela V, Aguilar-Hernández MM. Cellular signaling of the receptor for advanced glycation end products (RAGE). *Cell Signal.* 2013;25:2185-2197. doi: 10.1016/j.cellsig.2013.06.013.
91. Rubenstein DA, Maria Z, Yin W. Combined incubation of platelets and endothelial cells with glycated albumin: altered thrombogenic and inflammatory responses. *Diab Vasc Dis Res.* 2014;11:235-242. doi: 10.1177/1479164114531298.
92. Rubenstein DA, Morton BE, Yin W. The combined effects of sidestream smoke extracts and glycated serum albumin on endothelial cells and platelets. *Cardiovasc Diabetol.* 2010;9:28. doi: 10.1186/1475-2840-9-28.
93. Yamagishi SI, Matsui T, Takenaka K, Nakamura K, Takeuchi M, Inoue H. Pigment epithelium-derived factor (PEDF) prevents platelet activation and aggregation in diabetic rats by blocking deleterious effects of advanced glycation end products (AGEs). *Diabetes Metab Res Rev.* 2009;25:266-25271. doi: 10.1002/dmrr.906.
94. Maugeri N, Campana L, Gavina M, Covino C, De Metrio M, Panciroli C, et al. Activated platelets present high mobility group box 1 to neutrophils, inducing autophagy and promoting the extrusion of neutrophil extracellular traps. *J Thromb Haemost.* 2014;12:2074-2088. doi: 10.1111/jth.12710.
95. Koska J, Saremi A, Howell S, Bahn G, De Courten B, Ginsberg H, et al. Advanced glycation end products, oxidation products, and incident cardiovascular events in patients with type 2 diabetes. *Diabetes Care.* 2018;41(3):570-576. doi: 10.2337/dc17-1740.
96. Gerstein HC, Miller ME, Byington RP, Goff DC, Bigger JT, Buse JB, et al. Effects of intensive glucose lowering in type 2 diabetes. *N Engl J Med.* 2008;358(24):2545-2559. doi: 10.1056/NEJMoa0802743.
97. Saku K, Tahara N, Takaseya T, Otsuka H, Takagi K, Shojima T, et al. Pathological role of receptor for advanced glycation end products in calcified aortic valve stenosis. *J Am Heart Assoc.* 2020;9(13):e015261. doi: 10.1161/JAHA.119.015261.
98. McNair E, Qureshi M, Prasad K, Pearce C. Atherosclerosis and the hypercholesterolemic AGE-RAGE axis. *Int J Angiol.* 2016;25(02):110-116. doi: 10.1055/s-0035-1570754.
99. Reichert S, Triebert U, Santos AN, Hofmann B, Schaller H-G, Schlitt A, et al. Soluble form of receptor for advanced glycation end products and incidence of new cardiovascular events among patients with cardiovascular disease. *Atherosclerosis.* 2017;266:234-239. doi: 10.1016/j.atherosclerosis.2017.08.015.
100. Manganelli V, Truglia S, Capozzi A, Alessandri C, Riitano G, Spinelli FR, et al. Alarmin HMGB1, and soluble RAGE as new tools to evaluate the risk stratification in patients with the antiphospholipid syndrome. *Front Immunol.* 2019;10:460. doi: 10.3389/fimmu.2019.00460.
101. Al Rifai M, Schneider ALC, Alonso A, Maruthur N, Parrinello CM, Astor BC, et al. sRAGE, inflammation, and risk of atrial fibrillation: results from the Atherosclerosis Risk in Communities (ARIC) Study. *J Diabetes Compl.* 2015;29:180-185. doi: 10.1016/j.jdiacomp.2014.11.008.
102. Kaur N, Kishore L, Singh R. Attenuating diabetes: what really works? *Curr Diabetes Rev.* 2016;12:259e278. doi: 10.2174/1573399811666150826115410.
103. Pan D, Zhang D, Wu J, Chen C, Xu Z, Yang H, et al. A novel proteoglycan from *Ganoderma lucidum* fruiting bodies protects kidney function and ameliorates diabetic nephropathy via its antioxidant activity in C57BL/6 db/db mice. *Food Chem Toxicol.* 2014;63:111-118. doi: 10.1016/j.fct.2013.10.046.
104. Kishore L, Kaur N, Singh R. Distinct biomarkers for early diagnosis of diabetic nephropathy. *Curr Diabetes Rev.* 2017;13(6):598-605. doi: 10.2174/1573399812666161207123007.
105. Sasai Y, Iwakawa K, Yanagida K, Shen Y, Hosono T, Ariga T, et al. Advanced glycation endproducts stimulate renal epithelial cells to release chemokines that recruit macrophages, leading to renal fibrosis. *Biosci Biotechnol Biochem.* 2012;76(9):1741-1745. doi: 10.1271/bbb.120347.
106. Kaur N, Kishore L, Singh R. Therapeutic effect of *Linum usitatissimum* L. in STZ-nicotinamide induced diabetic nephropathy via inhibition of AGEs and oxidative stress. *J Food Sci Technol.* 2017;54:408-421. doi: 10.1007/s13197-016-2477-4.
107. Forbes JM, Coughlan MT, Cooper ME. Oxidative stress as a major culprit in kidney disease in diabetes. *Diabetes.* 2008;57(6):1446-1454. doi: 10.2337/db08-0057.
108. Stinghen AE, Massy ZA, Vlassara H, Striker GE, Boullier A. Uremic toxicity of advanced glycation end products in CKD. *J Am Soc Nephrol.* 2016;27(2):354-370. doi: 10.1681/ASN.2014101047.
109. Reiniger N, Lau K, McCalla D, Eby B, Cheng B, Lu Y, et al. Deletion of the receptor for advanced glycation end products reduces glomerulosclerosis and preserves renal function in the diabetic OVE26 mouse. *Diabetes.* 2010;59(8):2043-2054. doi: 10.2337/db09-1766.
110. Gerdemann A, Lemke HD, Nothdurft A, Heidland A, Münch G, Bahner U, et al. Low-molecular but not high-molecular advanced glycation end products (AGEs) are removed by high-flux dialysis. *Clin Nephrol.* 2000;54(4):276-283. PMID: 11076103.

111. Hellwig M, Geissler S, Matthes R, Peto A, Silow C, Brandsch M, et al. Transport of free and peptide-bound glycated amino acids: synthesis, transepithelial flux at Caco-2 cell monolayers, and interaction with apical membrane transport proteins. *Chembiochem*. 2011;12(8):1270-1279. doi: 10.1002/cbic.201000759.
112. Wihler C, Schäfer S, Schmid K, Deemer EK, Münch G, Bleich M, et al. Renal accumulation and clearance of advanced glycation end-products in type 2 diabetic nephropathy: effect of angiotensin-converting enzyme and vasopeptidase inhibition. *Diabetologia*. 2005;48(8):1645-1653. doi: 10.1007/s00125-005-1837-9.
113. Normand G, Lemoine S, Villien M, Le Bars D, Merida I, Irace Z, et al. AGE content of a protein load is responsible for renal performances: A pilot study. *Diabetes Care*. 2018;41:1292-1294. doi: 10.2337/dc18-0131.
114. Haraguchi R, Kohara Y, Matsubayashi K, Kitazawa R, Kitazawa S. New insights into the pathogenesis of diabetic nephropathy: Proximal renal tubules are primary target of oxidative stress in diabetic kidney. *Acta Histochem Cytochem*. 2020;53(2):21-31. doi: 10.1267/ahc.20008.
115. Perkins BA, Rabbani N, Weston A, Adaikalakoteswari A, Lee JA, Lovblom LE, et al. High fractional excretion of glycation adducts is associated with subsequent early decline in renal function in type 1 diabetes. *Sci Rep*. 2020;10(1):12709. doi: 10.1038/s41598-020-69350-y.
116. Vuong PM, Nguyen HT, Katano T, Matsumura S, Saito A, Yamada A, et al. Impaired peripheral nerve regeneration in type-2 diabetic mouse model. *Eur J Neurosci*. 2018;47:126-139. doi: 10.1111/ejn.13771.
117. Lee CC, Perkins BA, Kayaniyl S, Harris SB, Retnakaran R, Gerstein HC, et al. Peripheral neuropathy and nerve dysfunction in individuals at high risk for type 2 diabetes: The PROMISE Cohort. *Diabetes Care*. 2015;38(5):793-800. doi: 10.2337/dc14-2585.
118. Harati Y. Diabetic neuropathies: unanswered questions. *Neurol Clin*. 2007;25:303-317. doi: 10.1016/j.ncl.2007.01.002.
119. Leininger GM, Vincent AM, Feldman EL. The role of growth factors in diabetic peripheral neuropathy. *J Peripher Nerv Syst*. 2004;9:26-53. doi: 10.1111/j.1085-9489.2004.09105.x.
120. Du ZX, Zhang HY, Meng X, Guan Y, Wang HQ. Role of oxidative stress and intracellular glutathione in the sensitivity to apoptosis induced by proteasome inhibitor in thyroid cancer cells. *BMC Cancer*. 2009;9:56. doi: 10.1186/1471-2407-9-56.
121. Brownlee M. Negative consequences of glycation. *Metabolism*. 2000;49:9-13. doi: 10.1016/s0026-0495(00)80078-5.
122. Sekido H, Suzuki T, Jomori T, Takeuchi M, Yabe-Nishimura C, Yagihashi S. Reduced cell replication and induction of apoptosis by advanced glycation end products in rat Schwann cells. *Biochem Biophys Res Commun*. 2004;320(1):241-248. doi: 10.1016/j.bbrc.2004.05.159.
123. Kasajima H, Yamagishi S, Sugai S, Yagihashi N, Yagihashi S. Enhanced in situ expression of aldose reductase in peripheral nerve and renal glomeruli in diabetic patients. *Virchows Arch*. 2001;439(1):46-54. doi: 10.1007/s004280100444.
124. Vincent AM, Hinder LM, Pop-Busui R, Feldman EL. Hyperlipidemia: a new therapeutic target for diabetic neuropathy. *J Peripher Nerv Syst*. 2009;14:257-267. doi: 10.1111/j.1529-8027.2009.00237.x.
125. Al-Sofiani M, MacLeod S, Ghanim H, Stecker N, Hall J, Lippes H. Type 1 diabetes and hearing loss: Audiometric assessment and measurement of circulating levels of soluble receptor for advanced glycation end products. *Diabetes Metab Res Rev*. 2020;36:e3312. doi: 10.1002/dmrr.3312.
126. Zheng Y, He M, Congdon N. The worldwide epidemic of diabetic retinopathy. *Indian J Ophthalmol*. 2012;60(5):428-431. doi: 10.4103/0301-4738.100542.
127. Distefano LN, Garcia-Arumi J, Martinez-Castillo V, Boixadera A. Combination of anti-VEGF and laser photocoagulation for diabetic macular edema: a review. *J Ophthalmol*. 2017;2017:2407037. doi: 10.1155/2017/2407037.
128. Claramunt J. Diabetic retinopathy. *Revista Médica Clínica Las Condes*. 2009;20(5):670-679.
129. Cecilia O, Alberto CJ, José N, Germán CE, Ana Karen L, Miguel RL, et al. Oxidative stress as the main target in diabetic retinopathy pathophysiology. *J Diabetes Res*. 2019;2019:8562408. doi: 10.1155/2019/8562408.
130. Whitehead M, Wickremasinghe S, Osborne A, van Wijngaarden P, Martin KR. Diabetic retinopathy: a complex pathophysiology requiring novel therapeutic strategy. *Expert Opin Biol Ther*. 2018;18(12):1257-1270. doi: 10.1080/14712598.2018.1545836.
131. Behl T, Kotwani A. Exploring the various aspects of the pathological role of vascular endothelial growth factor (VEGF) in diabetic retinopathy. *Pharmacol Res*. 2015;99:137-148. doi: 10.1016/j.phrs.2015.05.013.
132. Schlotterer A, Kolibabka M, Lin J, Acunman K, Dietrich N, Sticht C, et al. Methylglyoxal induces retinopathy-type lesions in the absence of hyperglycemia: studies in a rat model. *FASEB J*. 2019;33(3):4141-4153. doi: 10.1096/fj.201801146RR.
133. Murakami T, Felinski EA, Antonetti DA. Occludin phosphorylation and ubiquitination regulate tight junction trafficking and vascular endothelial growth factor-induced permeability. *J Biol Chem*. 2009;284(31):21036-21046. doi: 10.1074/jbc.M109.016766.
134. Hendrick AM, Gibson MV, Kulshreshtha A. Diabetic Retinopathy. *Prim Care*. 2015;42(3):451-464. doi: 10.1016/j.pop.2015.05.005..
135. Bain SC, Klufas MA, Ho A, Matthews DR. Worsening of diabetic retinopathy with rapid improvement in systemic glucose control: a review. *Diabetes Obes Metab*. 2019;21(3):454-466. doi: 10.1111/dom.13538.
136. Giacco F, Brownlee M. Oxidative stress and diabetic complications. *Circ Res*. 2010;107(9):1058-1070. doi: 10.1161/CIRCRESAHA.110.223545.
137. Yan LJ. Redox imbalance stress in diabetes mellitus: role of the polyol pathway. *Animal Models Exp Med*. 2018;1(1):7-13. doi: 10.1002/ame2.12001.
138. Saxena R, Singh D, Saklani R, Gupta SK. Clinical biomarkers and molecular basis for optimized treatment of diabetic retinopathy: current status and future prospects. *Eye Brain*. 2016;8:1-13. doi: 10.2147/EB.S69185.
139. Obrosova I, Cao X, Greene DA, Stevens MJ. Diabetes-induced changes in lens antioxidant status, glucose utilization and energy metabolism: effect of DL- α -lipoic acid. *Diabetologia*. 1998;41(12):1442-1450. doi: 10.1007/s001250051090.

140. Das Evcimen N, King G. The role of protein kinase activation and the vascular complications of diabetes. *Pharmacol Res.* 2007;55(6):498-510. doi: 10.1016/j.phrs.2007.04.016.
141. Liu ZC, Yu EH, Liu W, Liu XC, Tang SB, Zhu BH. Translocation of protein kinase C δ contributes to the moderately high glucose-, but not hypoxia-induced proliferation in primary cultured human retinal endothelial cells. *Mol Med Rep.* 2014;9(5):1780-1786. doi: 10.3892/mmr.2014.2049.
142. Ramasamy R, Shekhtman A, Schmidt AM. The multiple faces of RAGE – opportunities for therapeutic intervention in aging and chronic disease. *Exp Opin Ther Targets.* 2016;20(4):431-446. doi: 10.1517/14728222.2016.1111873.
143. Homme RP, Singh M, Majumder A, George AK, Nair K, Sandhu HS, et al. Remodeling of retinal architecture in diabetic retinopathy: Disruption of ocular physiology and visual functions by inflammatory gene products and pyroptosis. *Front Physiol.* 2018;9:1268. doi: 10.3389/fphys.2018.01268.
144. El-Remessy AB, Al-Shabrawey M, Khalifa Y, Tsai NT, Caldwell RB, Liou GI. Neuroprotective and blood-retinal barrier-preserving effects of cannabidiol in experimental diabetes. *Am J Pathol.* 2006;168(1):235-244. doi: 10.2353/ajpath.2006.050500.
145. Tao D, Mai X, Zhang T, Mei Y. Association between the RAGE (receptor for advanced glycation end-products) - 374T/A gene polymorphism and diabetic retinopathy in T2DM. *Revista da Associação Médica Brasileira.* 2017;63(11):971-977. doi: 10.1590/1806-9282.63.11.971.
146. Kanda A, Dong Y, Noda K, Saito W, Ishida S. Advanced glycation endproducts link inflammatory cues to upregulation of galectin-1 in diabetic retinopathy. *Sci Rep.* 2017;7:16168. doi: 10.1038/s41598-017-16499-8.
147. Kandarakis SA, Piperi C, Topouzis F, Papavassiliou AG. Emerging role of advanced glycation end products (AGEs) in the pathobiology of eye diseases. *Prog Retin Eye Res.* 2014;42:85-102. doi: 10.1016/j.preteyeres.2014.05.002.
148. Shang F, Taylor A. Roles for the ubiquitin-proteasome pathway in protein quality control and signaling in the retina: implications in the pathogenesis of age-related macular degeneration. *Mol Aspects Med.* 2012;33:446-466. doi: 10.1016/j.mam.2012.04.001.
149. Malik NS, Moss SJ, Ahmed N, Furth AJ, Wall RS, Meek KM. Aging of the human corneal stroma: structural and biochemical changes. *Biochim Biophys Acta.* 1992;20:222-228. doi: 10.1016/0925-4439(92)90041-k.
150. Brummer G, Littlechild S, McCall S, Zhang Y, Conrad GW. The role of non-enzymatic glycation and carbonyls in collagen cross-linking for the treatment of keratoconus. *Invest Ophthalmol Vis Sci.* 2011;52:6363-6369. doi: 10.1167/iovs.11-7585.
151. Shi L, Chen H, Yu X, Wu X. Advanced glycation end products delay cornea epithelial wound healing through reactive oxygen species generation. *Mol Cell Biochem.* 2013;383:253-259. doi: 10.1007/s11010-013-1773-9.
152. Shi L, Yu X, Yang H, Wu X. Advanced glycation end products induce human corneal epithelial cells apoptosis through the generation of reactive oxygen species and activation of JNK and p38 MAPK pathways. *PLoS One.* 2013;8:e66781. doi: 10.1371/journal.pone.0066781.
153. Zou C, Wang S, Huang F, Zhang YA. Advanced glycation end products and ultrastructural changes in corneas of long-term streptozotocin-induced diabetic monkeys. *Cornea.* 2012;31:1455-1459. doi: 10.1097/ICO.0b013e3182490907.
154. Nagaraj RH, Kern TS, Sell DR, Fogarty J, Engerman RL, Monnier VM. Evidence of a glycemic threshold for the formation of pentosidine in diabetic dog lens but not I collagen. *Diabetes.* 1996;45:587-594. doi: 10.2337/diab.45.5.587.
155. Swamy-Mruthinti S, Shaw SM, Zhao HR, Green K, Abraham EC. Evidence of glycemic threshold for the development of cataracts in diabetic rats. *Curr Eye Res.* 1999;18:423-429. doi: 10.1076/ceyr.18.6.423.5271.
156. Raghavan C, Smuda M, Smith A, Howell S, Smith D, Singh A, et al. AGEs in human lens capsule promote the TGF β 2-mediated epithelial to mesenchymal transition of lens epithelial cells implications for posterior capsule opacification. *Aging Cell.* 2016;15(3):465-476. doi: 10.1111/acel.12450.
157. Kim J, Kim OS, Kim CS, Sohn E, Jo K, Kim JS. Accumulation of argpyrimidine a methylglyoxal-derived advanced glycation end product increases apoptosis of lens epithelial cells both in vitro and in vivo. *Exp Mol Med.* 2012;44:167-175. doi: 10.3858/emmm.2012.44.2.012.
158. Sebag J, Buckingham B, Charles MA, Reiser K. Biochemical abnormalities in vitreous of humans with proliferative diabetic retinopathy. *Arch Ophthalmol.* 1992;110:1472-1476. doi: 10.1001/archophth.1992.01080220134035.
159. Lee OT, Good SD, Lamy R, Kudisch M, Stewart JM. Advanced glycation end product accumulation reduces vitreous permeability. *Invest Ophthalmol Vis Sci.* 2015;56:2892-2897. doi: 10.1167/iovs.14-15840.
160. Terai N, Spoerl E, Hausteiner M, Hornykewycz K, Haentzschel J, Pillunat LE. Diabetes mellitus affects the biomechanical properties of the optic nerve head in the rat. *Ophthalm Res.* 2012;47:189-194. doi: 10.1159/000331990.
161. Albon J, Karwatowski WS, Easty DL, Sims TJ, Duance VC. Age-related changes in the non-collagenous components of the extracellular matrix of the human lamina cribrosa. *Br J Ophthalmol.* 2000;84:311-317. doi: 10.1136/bjo.84.3.311.
162. Shanbhogue VV, Mitchell DM, Rosen CJ, Bouxsein ML. Type 2 diabetes and the skeleton: new insights into sweet bones. *Lancet Diabetes Endocrinol.* 2016;4(2):159-173. doi: 10.1016/S2213-8587(15)00283-1.
163. Farr JN, Khosla S. Determinants of bone strength and quality in diabetes mellitus in humans. *Bone.* 2016;82:28-34. doi: 10.1016/j.bone.2015.07.027.
164. Ott C, Jacobs K, Haucke E, Navarrete Santos A, Grune T, Simm A. Role of advanced glycation end products in cellular signaling. *Redox Biol.* 2014;2:411-429. doi: 10.1016/j.redox.2013.12.016.
165. Zhou Z, Xiong WC. RAGE and its ligands in bone metabolism. *Front Biosci.* 2011;3:768-776. doi: 10.2741/s185.
166. Chen X, Wang Z, Duan N, Zhu G, Schwarz EM, Xie C. Osteoblast-osteoclast interactions. *Connect Tissue Res.* 2018;59(2):99-107. doi: 10.1080/03008207.2017.1290085.
167. Oren TW, Botolin S, Williams A, Bucknell A, King KB. Arthroplasty in veterans: analysis of cartilage, bone, serum, and synovial fluid reveals differences and similarities in osteoarthritis with and without comorbid diabetes. *J Rehabil*

- Res Dev.* 2011;48(10):1195-1210. doi: 10.1682/jrdd.2010.09.0186.
168. Franke S, Siggelkow H, Wolf G, Hein G. Advanced glycation endproducts influence the mRNA expression of RAGE, RANKL and various osteoblastic genes in human osteoblasts. *Arch Physiol Biochem.* 2007;113(3):154-161. doi: 10.1080/13813450701602523.
 169. Notsu M, Yamaguchi T, Okazaki K, Tanaka K, Ogawa N, Kanazawa I, et al. Advanced glycation end product 3 (AGE3) suppresses the mineralization of mouse stromal ST2 cells and human mesenchymal stem cells by increasing TGF- β expression and secretion. *Endocrinology.* 2014;155(7):2402-2410. doi: 10.1210/en.2013-1818.
 170. Okazaki K, Yamaguchi T, Tanaka K, Notsu M, Ogawa N, Yano S, et al. Advanced glycation end products (AGEs), but not high glucose, inhibit the osteoblastic differentiation of mouse stromal ST2 cells through the suppression of osterix expression, and inhibit cell growth and increasing cell apoptosis. *Calcif Tissue Int.* 2012;91(4):286-296. doi: 10.1007/s00223-012-9641-2.
 171. Meng HZ, Zhang WL, Liu F, Yang MW. Advanced glycation end-products affect osteoblast proliferation and function by modulating autophagy via the receptor of advanced glycation end products/ Raf protein/mitogen-activated protein kinase/extracellular signal-regulated kinase/ extracellular signal-regulated kinase (RAGE/Raf/MEK/ERK) pathway. *J Biol Chem.* 2015;290(47):28189-28199. doi: 10.1074/jbc.M115.669499.
 172. Khosravi R, Sodek KL, Faibish M, Trackman PC. Collagen advanced glycation inhibits its discoidin domain receptor 2 (DDR2)-mediated induction of lysyl oxidase in osteoblasts. *Bone.* 2014;58:33-41. doi: 10.1016/j.bone.2013.10.001.
 173. Li W, Ling W, Teng X, Quan C, Cai S, Hu S. Effect of advanced glycation end products, extracellular matrix metalloproteinase inducer, and matrix metalloproteinases on type-I collagen metabolism. *Biomed Rep.* 2016; 4(6):691-693. doi: 10.3892/br.2016.641.
 174. Owen WF, Hou FF, Stuart RO, Kay J, Boyce J, Chertow GM, et al. b2-Microglobulin modified with advanced glycation end products modulates collagen synthesis by human fibroblasts. *Kidney Int.* 1998;53(5):1365-1373. doi: 10.1046/j.1523-1755.1998.00882.x.
 175. Kumar Pasupulati A, Chitra PS, Reddy GB. Advanced glycation end products mediated cellular and molecular events in the pathology of diabetic nephropathy. *Biomol Concepts.* 2016;7(5-6):293-309. doi: 10.1515/bmc-2016-0021.
 176. Rabelo GD, Roux JP, Portero-Muzy N, Gineyts E, Chapurlat R, Chavassieux P. Cortical fractal analysis and collagen crosslinks content in the femoral neck after osteoporotic fracture in postmenopausal women: comparison with osteoarthritis. *Calcif Tissue Int.* 2018;102(6):644-650. doi: 10.1007/s00223-017-0378-9.
 177. Tamaki J, Kouda K, Fujita Y, Iki M, Yura A, Miura M, et al. Ratio of endogenous secretory receptor for advanced glycation end products to pentosidine predicts fractures in men. *J Clin Endocrinol Metab.* 2018;103(1):85-94. doi: 10.1210/jc.2017-00929.
 178. Vaculik J, Braun M, Dungal P, Pavelka K, Stepan JJ. Serum and bone pentosidine in patients with low impact hip fractures and patients with advanced osteoarthritis. *BMC Musculoskelet Disord.* 2016;17(1):308. doi: 10.1186/s12891-016-1168-7.
 179. Farlay D, Armas LA, Gineyts E, Akhter MP, Recker RR, Boivin G. Non-enzymatic glycation and degree of mineralization are higher in bone from fractured patients with type 1 diabetes mellitus. *J Bone Miner Res.* 2016;31(1):190-195. doi: 10.1002/jbmr.2607.
 180. Wang Z, Zhang J, Chen L, Li J, Zhang H, Guo X. Glycine Suppresses AGE/RAGE Signaling Pathway and Subsequent Oxidative Stress by Restoring Glo1 Function in the Aorta of Diabetic Rats and HUVECs. *Oxid Med Cell Longev.* 2019; 2019:4628962. doi: 10.1155/2019/4628962.
 181. Adeshara KA, Bangar NS, Doshi PR, Diwan A, Tupe RS. Action of metformin therapy against advanced glycation, oxidative stress, and inflammation in type 2 diabetes patients: 3 months of follow-up study. *Diabetes Metab Syndr Clin Res Rev.* 2020;14(5):1449-1458. doi: 10.1016/j.dsx.2020.07.036.
 182. Li H, Chen J, Li B, Fang X. The protective effects of dulaglutide against advanced glycation end products (AGEs)-induced degradation of type II collagen and aggrecan in human SW1353 chondrocytes. *Chem Biol Interact.* 2020;322:108968. doi: 10.1016/j.cbi.2020.108968.
 183. Omidian M, Djalali M, Javanbakht MH, Eshraghian MR, Abshirini M, Omidian P, et al. Effects of vitamin D supplementation on advanced glycation end products signaling pathway in T2DM patients: a randomized, placebo-controlled, double blind clinical trial. *Diabetol Metab Syndr.* 2019;11:86. doi: 10.1186/s13098-019-0479-x.